Sepsis ICD Coding Validation Study

Validation of ICD coding methods in estimating sepsis epidemiology: A prospective, observational cohort study

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STATEMENT OF COMPLIANCE

This document is a protocol for a clinical research study. The study will be conducted in compliance with all stipulations of this protocol, the conditions of ethics committee approval, the NHMRC National Statement on Ethical Conduct in Human Research (2007) and the Note for Guidance on Good Clinical Practice (CPMP/ICH-135/95) annotated with Therapeutic goods Administration comments.



1 Synopsis

Background: In 2017, The World Health Organization recognised sepsis as a global health priority.¹ However, estimation of true disease burden of sepsis remains difficult in lieu of lack of diagnostic tests and inconsistencies in the International Classification of Diseases (ICD) coding methodologies. We aim to use modification of IHME method to develop a reliable and reproducible estimate of the true sepsis burden in Australia using calibrated ICD coding methods.

Aim: To estimate the predictive value of sepsis ICD coding method using medical chart reviews and to compare the predictive values of modified-IHME and Angus ICD coding methods.

Study design: Prospective, observational, multicentre cohort study

Study population: Adult patients (aged 16 years old or more) admitted from the Emergency Department (ED) with an order of culture of any body fluid plus oral or intravenous administration of an antibiotic plus presence of ≥ 2 qSOFA score.

Methods: All ED admissions from 05 South West Sydney Local health District (SWSLHD) hospitals in NSW will be screened by a member of the research team to select 500 adult patients (aged 16 years old or more) using the study inclusion criteria. The two reviewers will review medical charts of the selected patients to determine if they meet the SEPSIS-3 criteria. The clinical diagnosis will be used to determine the predictive value of ICD coding. An analysis of initial 100 patients will be done to check if about 50% patients have clinical sepsis in the selected cohort and if required, inclusion criteria will be modified accordingly.

Data collection: Demographic and clinical characteristics of the selected cases will be recorded at the time of admission. At 96 hours after hospital admission, medical chart review will be done to identify clinical sepsis cases as per SEPSIS-3 criteria. For such cases the following additional information will be collected: onset of sepsis episode, site of infection, causative organism, time to antibiotic, type of organ dysfunction and presence of septic shock. On day 60 after hospital discharge, the assigned ICD codes and possibly death certificate data for patients who will die, will be obtained from the medical record department to apply Angus and modified-IHME methods.

Outcome: The findings from this study will provide valuable insights to calibrate ICD coding methods to give accurate and repeatable estimates of the burden of sepsis in SWSLHD in NSW which can be applied in other Australian states and territories and nationally.

2 Administrative Information

2.1 Investigator & participating Institutions

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Dr Paul M Middleton, Deputy Director, Emergency Medicine, Liverpool Hospital

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Dr Matthew Smith, Director of Emergency Medicine, Bankstown Hospital

Dr Kavita Shetty, ICU Specialist, Fairfield Hospital

Note: More sites and site PIs may be added as necessary to achieve the study sample size

2.2 Study management committee

<u>Dr Ashwani Kumar:</u> Research Associate, PhD Candidate, Critical Care Division, The George Institute for Global Health, NSW, Australia

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3 General Information

3.1 Title

Validation of ICD coding methods in estimating sepsis epidemiology: A prospective, multicentre cohort study.

3.2 Short Title

Sepsis ICD coding validation study.

4 Background and Rationale for Project

Sepsis is a major health problem and is associated with significant mortality and morbidity.¹⁻ ³ However, the burden of sepsis remains poorly understood both globally and in Australia, primarily because there is no definitive diagnostic test. As a result, use of administrative data such as ICD codes remains the only practical means to estimate sepsis epidemiology on a large scale.⁴⁻⁷ ICD codes are the diagnostic classification standard proposed by WHO for all diseases disorders, injuries and other related health conditions for clinical and research purposes. These codes are assigned after a patient gets discharged from the hospital based on the primary and secondary diagnosis by designated hospital staff called Medical Coders. Several authors have used ICD coding methods to study the epidemiology of sepsis, amongst which Angus⁸ and Martin⁹ criteria are the most well-known and highly cited. However, currently there is no consensus regarding which ICD coding method should be used to define sepsis in administrative data and thereby estimate the true disease burden of sepsis.

The two predominant strategies to identify sepsis using ICD codes are 1) using specific ICD codes for sepsis, severe sepsis and septic shock, known as the 'explicit method' or 2) the presence of the ICD codes for infection and organ dysfunction during the same hospital episode, known as the "implicit" method. However, both strategies have some limitations. Explicit sepsis codes are only applied if sepsis or septic shock is documented in the clinical notes by the clinicians, which is not a common practice. The Implicit method comes with its own set of problems as various combination of infection and diagnosis codes have been proposed to identify sepsis, each suggesting different sepsis rates. Moreover, documentation of infection and organ dysfunction codes at discharge do not confirm if organ dysfunction was induced by infection or even existed at the same time during the hospital episode. Another issue is significant difference in the sepsis ICD coding practices, its documentation and selection of specific codes not just amongst different countries but within the same country or even same state. In a recent high-quality study in the USA using the Angus criteria, the explicit method underestimated sepsis incidence by approximately 50% whereas the implicit method overestimated incidence by 100%. These limitations result in estimates of the sepsis incidence varying up to four-fold depending on the method used.¹⁰ Moreover, most previous studies which estimated the predictive value of ICD coding were done using retrospective medical chart review. This design has an inherent limitation that information captured in the medical charts cannot be verified. Another common design is the analysis of administrative datasets which leverages the efficiency of large pre-existing data but is limited by variations in physician documentation and hospital coding practices.

Recently, the Institute for Health Metrics and Evaluation (IHME), an independent population health research centre at the University of Washington and WHO Global Burden of Disease (GBD) group, proposed a new ICD coding method to estimate the global sepsis disease burden. This method, which also classifies sepsis cases as Explicit or Implicit, differs from earlier estimates not only in its scope but also in using death certificate data, rather than hospitalization data.¹¹ However, it has not been prospectively validated. We aim to use modification of IHME method to develop a reliable and reproducible estimate of the burden of sepsis in SWSLHD in NSW, Australia by calibrating ICD coding of hospitalisation data using prospective medical chart review.

4.1 Aims and Objectives

4.1.1 Primary objectives

To estimate the predictive value of the modified IHME/GBD ICD coding method using prospective medical chart review.

4.1.2 Secondary objective

To compare the predictive value of modified IHME/GBD and Angus coding methods in estimating sepsis epidemiology.

5 Study Design

A prospective, observational, multicentre cohort study.

5.1 Inclusion criteria

- Adult patients (aged 16 years old or more) admitted from the Emergency Department (ED) with an expected duration of hospital stay >24 hrs
- An order of culture of body fluid plus oral or intravenous administration of an antibiotic
- Presence of two of the following criteria (qSOFA score)
 - Altered mental status (GCS <15)
 - Respiratory rate <a>22/min
 - Systolic blood pressure <100 mmHg

5.2 Data source

Data will be extracted from patient's clinical notes and medical records during the hospital stay.

5.3 Data collection

Demographic and clinical characteristics of the selected patients will be documented over their hospital admission period:

At admission

- MRN
- Age
- Sex
- qSOFA score
 - Respiratory rate
 - Systolic blood pressure
 - Glasgow Coma Score
- Baseline SOFA score, if available
- Order and/or culture of body fluid
- Order and/or administration of an antibiotic
- Co-morbidities

Follow-up at 96 hours

- ICU admission: Yes/No
- Total SOFA score

If total SOFA score is not documented in clinical notes, individual SOFA component scores will be calculated using SOFA worksheet (Appendix C).

For cases identified as having sepsis, following additional information will be collected:

- Date of sepsis episode
- Type of organ dysfunction
- Presence of septic shock
 - Use of a vasopressor to maintain Mean Arterial Pressure >65 mmHg, PLUS
 - Lactate > 2 mmol/dL (18 mg/dL)

Day 60 (post admission)

Following data will be obtained from the hospital medical record department (MRD):

- Assigned ICD codes
 - Primary diagnosis
 - Secondary diagnosis(es)
- Type of admission
 - Surgical
 - Elective
 - Emergency

- o Medical
- Death Certificate data (for patients who will die)
- Time to antibiotic
- Site of infection
- Causative organism
- In-hospital LOS
- In-hospital mortality

Note: Patients who didn't meet clinical sepsis criteria at 96 hour follow up will be again evaluated at day 60 follow up if they met sepsis criteria between 96 hour and their discharge

If the same patient comes back to hospital during the study period, it will be considered as a separate admission.

6 Methods

All ED admissions from SouthWest Sydney Local Health District (SWSLHD) hospitals (Liverpool, Bankstown, Campbelltown, Bowral and Fairfield) in New South Wales will be screened by a member of the research team by applying the study inclusion criteria. Sites may choose to do the screening on 15-20 days in a month. More sites can be added, as required.

At 96 hrs after the admission, selected cases will be independently reviewed by two members of the research team to determine if they meet the clinical sepsis criteria. In case of disagreement between the two reviewers, the treating physician will be asked to make the judgement.

Clinical sepsis criteria (SEPSIS-3):

1. Presumed serious infection:

- Order or documented administration of oral or parenteral antibiotics, PLUS
- Order or documented sampling of body fluid cultures (blood, urine, cerebrospinal fluid, peritoneal, etc.) in a specified period[#].

[#]If culture is obtained first, antibiotic must be administered within the next 72 hours, whereas if antibiotic is administered first, culture is required within the next 24 hours.

AND

2. Acute organ dysfunction

Infection-related increase in Sequential Organ Failure Assessment (SOFA) score of \geq 2 from baseline

Note:

Baseline SOFA scores will be determined by the reviewer from the available data in the medical chart, estimating the degree of organ dysfunction prior to infection onset (if infection occurred during hospitalization) or at baseline health (if infection was present-on-admission). If no baseline SOFA score is available, then it should be assumed as '0'.

Last data collection will include principal and secondary diagnosis ICD codes along with some additional information which will be obtained from the MRD of each hospital. It will be determined by completion of coding which we expect to get completed by 60 days after

hospital discharge. Wherever possible, death certificate data will be collected for patients who will die.

Angus (Appendix A) and modified IHME/GBD (Appendix B) sepsis methods will be applied to identify database sepsis diagnosis.

Explicit sepsis cases will be those with an ICD code explicitly referencing sepsis (Appendix B.1) listed as a main cause of death or secondary admission diagnosis while implicit sepsis cases will be those with both an "infection code" listed as the underlying cause of death or primary admission diagnosis and an "organ dysfunction code" listed as a main cause of death or secondary admission diagnosis. If a case will meet criteria for both case definitions, it will be classified as explicit. Implicit sepsis cases will be identified using both Angus and modified-IHME criteria.

The predictive value of both ICD coding methods will be calculated using clinical diagnosis of sepsis as the gold standard.

Following table will be created for both Angus and modified-IHME ICD coding methods:

	Clinical sepsis (n)	
Database sepsis (n)	Yes	No
Yes	True Positive (TP)	False Positive (FP)
No	False Negative (FN)	True Negative (TN)

Predictive value parameters will be calculated as follows:

- Positive Predictive Value (PPV) = TP/(TP + FP) x 100
- Negative Predictive Value (NPV) = TN/(TN + FN) x 100
- Sensitivity = TP/TP+FN
- Specificity = TN/TN+FP

7 Safety Monitoring

As this is an observational study using data collected as part of routine clinical practice, no safety monitoring will be undertaken.

8 Statistical Analysis Plan

8.1 Sample size

Sample size calculation is based on considerations for the width of a two-sided 95% confidence interval (CI) of reported sensitivity of explicit sepsis ICD-10 codes between 7-16%.¹²⁻¹⁴ For optimal statistical analysis, 50% of the enrolled patients should have the diagnosis of clinical sepsis in the selected patient cohort. For this reason and low incidence of sepsis in general population, inclusion criteria of presumed infection plus positive qSOFA will be used to select a patient's cohort with increased predicted risk of developing sepsis. A previous study in patients with diagnosed infection reported the proportion of Sepsis-3 positive cases as 32%¹⁵ whereas a recently published Chinese study reported the sensitivity of qSOFA as 50.4% in predicting sepsis diagnosis in patients with infection.¹⁶ Hence, we expect about 50% clinical sepsis cases (using SEPSIS-3 criteria) in our selected patient cohort and a sample size of 500 should provide about 250 cases of true sepsis cases. In lieu of limited

overall evidence and none from Australia on the predictive ability of our proposed eligibility criteria, we will conduct an initial analysis of first 100 patients to check the proportion of the clinical sepsis in our cohort. Based on the interim analysis results, study eligibility criteria will be modified, if required.

8.2 Data analysis

A descriptive analysis of the demographic and clinical characteristics will be presented. Continuous, normally distributed data will be presented as means with standard deviations (SDs), non-normally distributed data as medians with interquartile ranges (IQRs). Categorical data will be presented as numbers and proportions. Estimates of incidence will be presented with 95% confidence intervals (CIs).

9 Data Management

The principal means of data collection and data processing will be via online data entry performed by a member of the research team.

9.1 Site monitoring

No site monitoring will be performed.

9.2 Data recording and document retention

Each participating site will enter de-identified data into electronic CRF (eCRF). Each site will access the system through an individual password protected account and will only have access to their own site's data. Data will be stored in the Principal Investigator's office in electronic form on a database used specifically for the project.

All data collected by the study will be kept for a minimum of 7 years, or as otherwise required by regulatory authorities. Any paper data will be archived and stored in a secure facility.

9.3 Data quality assurance

Data entered by each site will be password-protected and the ability to access or change data prior to locking of the database will be restricted to that site and designated member of the research team. Once initial data collection has been completed, missing data and implausible values will be identified using predetermined objective criteria, and queries resolved through direct communication with sites if required. Treatment of outliers and missing data will be in accordance with a statistical analysis plan.

10 Outcome

The proposed study presents an opportunity to develop better ICD coding methods and to calibrate them using prospective clinical diagnosis to give an accurate and reproducible estimate of burden of sepsis in NSW, which can be applied to other Australian states and territories and nationally. Though, determining predictive ability of qSOFA for sepsis diagnosis is not a study objective, this study will provide additional valuable insights about the same as only a few studies have evaluated qSOFA as the predictor of sepsis diagnosis.

11 Funding

This study is currently being funded by internal sources from The George Institute for Global Health. Future funding will be sought via suitable grants.

12 Project Timeline

Anticipated timelines:

Date	Project Milestone
Jul 2019 – Jan 2020	Finalise Protocol
Nov 2019 – Mar 2020	Finalise list of participating hospitals in NSW
Feb 2020 – Jul 2020	Ethics submission & approval for participating hospitals
Jul - Nov 2020	Preparation of study documents, development of eCRF
Nov 2020 - Jan 2021	Expected enrolment start date
Jul 2022 – Sep 2022	Expected enrolment end date
Jan - Mar 2022	Data entry completed
Apr – Jun 2022	Data analysis
Jul - Nov 2022	Preparation of manuscript
Dec 2022 - Mar 2023	Presentation and publication of results

13 Ethical considerations

This is a low/negligible risk observational study as it is completely non-invasive in that it uses demographic, physiological and patient stay data which is measured routinely as part of clinical care. There is no change in the use of these data, nor is there data collected which is extra to that used in routine patient assessment and management. There are no interventions in the study and patient management will not be influenced in any way. Gaining informed consent in the ED setting has been the subject of much debate, and in the circumstances of observational nature of this study, the risk to the individual patient is extremely low, and essentially translates to those risks surrounding privacy principles. The NHMRC National Statement (section 2.3.6 (a)) requires that the research 'carries no more than low risk' and that the 'only foreseeable risk is one of discomfort'.

Alternatives to prospective informed consent include proxy consent, a waiver from individual participant consent and retrospective or deferred consent. There is evidence in proxy consent that the proxy may demonstrate poor agreement with the wishes of the participant; also, proxy consent may not be available in the timely manner necessary for interventions with a narrow therapeutic window. Deferred consent is used when it is not possible to obtain prospective informed consent from the participant or proxy, and consent is obtained from the participant as soon as is feasible after the intervention. However, not only does deferred consent run the risk of loss to follow up of patients enrolled in the study, but particularly in the emergency setting also involves attempts to contact the patient after discharge, a process which may cause unnecessary psychological distress and anxiety. A waiver of individual consent has been suggested as a solution to many 'low-risk' investigations in emergency

medicine, allowing consideration of factors such as minimal or no risk to the patient or staff, and the mandate to produce sound research-based evidence for time-critical interventions.

In the current study, there is no clinical risk associated for participants as the study is observational in nature and has no potential to interfere with standard treatment. There is no risk to the rights, privacy or professional reputation of carers, health professionals and/or institutions as the study solely concerns with analysis of clinical data collected as part of standard clinical care. Moreover, there won't be any clinical interaction between study participants and the research team as data will be captured from patients' clinical notes and medical records. Given the observational nature of the study and no perceived risks to patient health or safety, ethical approval will be sought with a waiver of consent.

14 Regulatory Requirements

This study will be conducted in accordance with the ICH and GCP principles.

15 Publication Policy

Data collected in the study will be presented at Intensive care conferences and other forums as appropriate. Data will subsequently be used for scientific publications in academic journals. It may also provide support for research applications.

16 Trial Registration

ANZCTR registration no. ACTRN12621000333819.

17 References

- Stopping Sepsis: A National Action Plan [Internet]. 2017 p. 3. Available from: <u>https://www.georgeinstitute.org.au/sites/default/files/documents/stopping-sepsis-national-action-plan.pdf</u>
- Torio CM, Moore BJ. National inpatient hospital costs: the most expensive conditions by payer, 2013. Available from: <u>https://www.hcup-us.ahrq.gov/reports/statbriefs/sb204-Most-Expensive-Hospital-Conditions.jsp</u>
- 3. <u>http://apps.who.int/gb/ebwha/pdf_files/WHA70/A70_R7-en.pdf</u>
- 4. Gobatto, A., Besen, B. and Azevedo, L. (2017). How Can We Estimate Sepsis Incidence and Mortality? SHOCK, 47, p.6-11.
- 5. Jolley, R., Sawka, K., Yergens, D., Quan, H., Jetté, N. and Doig, C. (2015). Validity of administrative data in recording sepsis: a systematic review. Critical Care, 19(1).
- 6. Klompas, M. and Rhee, C. (2016). Sepsis and the theory of relativity: measuring a moving target with a moving measuring stick. Critical Care, 20(1).
- 7. Tsertsvadze, A., Royle, P., Seedat, F., Cooper, J., Crosby, R. and McCarthy, N. (2016). Community-onset sepsis and its public health burden: a systematic review. Systematic Reviews, 5(1).
- 8. Angus, D., Linde-Zwirble, W., Lidicker, J., Clermont, G., Carcillo, J., & Pinsky, M. (2001). Epidemiology of severe sepsis in the United States: Analysis of incidence, outcome, and associated costs of care. Critical Care Medicine, 29(7), 1303-1310.
- 9. Martin, G., Mannino, D., Eaton, S., & Moss, M. (2003). The Epidemiology of Sepsis in the United States from 1979 through 2000. New England Journal of Medicine, 348(16), 1546-1554.
- Rhee, C., Dantes, R., Epstein, L., Murphy, D., Seymour, C., Iwashyna, T., Kadri, S., Angus, D., Danner, R., Fiore, A., Jernigan, J., Martin, G., Septimus, E., Warren, D., Karcz, A., Chan, C., Menchaca, J., Wang, R., Gruber, S. and Klompas, M. (2017). Incidence and Trends of Sepsis in US Hospitals Using Clinical vs Claims Data, 2009-2014. JAMA, 318(13), 1241

- Rudd, K., Johnson, S., Agesa, K., Shackelford, K., Tsoi, D., & Kievlan, D. et al. (2020). Global, regional, and national sepsis incidence and mortality, 1990–2017: analysis for the Global Burden of Disease Study. The Lancet, 395(10219), 200-211
- 12. Angus DC, Linde-Zwirble WT, Lidicker J, Clermont G, Carcillo J, Pinsky MR. (2001). Epidemiology of severe sepsis in the United States: analysis of incidence, outcome, and associated costs of care. Critical care medicine.; 29(7):1303-10
- Ibrahim I, Jacobs IG, Webb SAR, Finn F. (2012). Accuracy of International classification of diseases, 10th revision codes for identifying severe sepsis in patients admitted from the emergency department Crit Care Resusc, 14: 112–118
- 14. Iwashyna, T., Odden, A., Rohde, J., Bonham, C., Kuhn, L., & Malani, P. et al. (2014). Identifying Patients with Severe Sepsis Using Administrative Claims. Medical Care, 52(6), e39-e43.
- 15. Mellhammar, L., Wullt, S., Lindberg, Å., Lanbeck, P., Christensson, B., & Linder, A. (2016). Sepsis Incidence: A Population-Based Study. Open Forum Infectious Diseases, 3(4), ofw207.
- 16. Tian, H., Zhou, J., Weng, L., Hu, X., Peng, J., & Wang, C. et al. (2019). Accuracy of qSOFA for the diagnosis of sepsis-3: a secondary analysis of a population-based cohort study. Journal Of Thoracic Disease, 11(5), 2034-2042.

18 Appendices

Appendix A: Angus Sepsis Definition

ICD-10-AM translation of infection codes used by	y Angus et al ²⁵
A00 Cholera	A51 Early syphilis
A01 Typhoid and paratyphoid fevers	A52 Late syphilis
A02 Other salmonella infections	A53 Other and unspecified syphilis
A03 Shigellosis	A54 Gonococcal infection
A04 Other bacterial intestinal infections	A65 Nonvenereal syphilis
A05 Other bacterial food-borne intoxications	A66 Yaws
08 Viral and other specified intestinal infections	A67 Pinta (carate)
09 Diarrhea and gastroenteritis, presumed infectious origin	A69 Other spirochetal infections
15 Respiratory tuberculosis, bacteriologically and histologically	B35 Dermatophytosis
onfirmed	B36 Other superficial mycoses
16 Respiratory tuberculosis, confirmed bacteriologically or	B37 Candidiasis
istologically	B38 Coccidioidomycosis
17 Tuberculosis of nervous system	B39 Histoplasmosis
18 Tuberculosis of other organs	B40 Blastomycosis
19 Miliary tuberculosis	B41 Paracoccidioidomycosis
N20 Plague	B42 Sporotrichosis
.21 Tularemia	B43 Chromomycosis and phaeomycotic abscess
22 Anthrax	B44 Aspergillosis
23 Brucellosis	B45 Cryptococcosis
24 Glanders and melioidosis	B46 Zygomycosis
25 Rat-bite fevers	B47 Mycetoma
A27 Leptospirosis	B48 Other mycoses, not elsewhere classified
28 Other zoonotic bacterial diseases, not elsewhere classified	B49 Unspecified mycosis
A30 Leprosy (Hansen's disease)	G00 Bacterial meningitis, not elsewhere classified
A31 Infection due to other mycobacteria	G01 Meningitis in bacterial diseases classified elsewhere
A32 Listeriosis	G02 Meningitis in other infectious and parasitic disease
A33 Tetanus neonatorum	classified elsewhere
34 Obstetrical tetanus	G03 Meningitis due to other and unspecified causes
N35 Other tetanus	G04 Encephalitis, myelitis and encephalomyelitis
N36 Diphtheria	G05 Encephalitis, myelitis and encephalomyelitis in diseas
N37 Whooping cough	classified elsewhere
A38 Scarlet fever	G06 Intracranial and intraspinal abscess and granuloma
A39 Meningococcal infection	G07 Intracranial and intraspinal abscess and granuloma
A40 Streptococcal sepsis	diseases classified elsewhere
41 Other sepsis	G08 Intracranial and intraspinal phlebitis and thrombophlebi
42 Actinomycosis	G09 Sequelae of inflammatory diseases of central nervo
43 Nocardiosis	system
A44 Bartonellosis	I30 Acute pericarditis
A46 Erysipelas	I33 Acute and subacute endocarditis
A48 Other bacterial diseases, elsewhere classified	I80 Phlebitis and thrombophlebitis
A50 Congenital syphilis	J01 Acute sinusitis
A49 Bacterial infection of unspecified site	J02 Acute pharyngitis

103 Acute tonsillitis J04 Acute laryngitis and tracheitis J05 Acute obstructive laryngitis (croup) and epiglottitis J06 Acute upper respiratory infections of multiple and unspecified sites J13 Pneumonia due to Streptococcus pneumoniae J14 Pneumonia due to Haemophilus influenzae J15 Bacterial pneumonia, not elsewhere classified J16 Pneumonia due to other infectious organisms, not elsewhere classified J17 Pneumonia in diseases classified elsewhere J18 Pneumonia, organism unspecified J44.0 Chronic obstructive pulmonary disease with acute lower respiratory infection J44.1 Chronic obstructive pulmonary disease with acute exacerbation, unspecified J47 Bronchiectasis J85 Abscess of lung and mediastinum J86 Pyothorax K35 Acute appendicitis K36 Other appendicitis K37 Unspecified appendicitis K57.02 Diverticulitis of small intestine with perforation and abscess, no hemorrhage K57.03 Diverticulitis of small intestine with perforation and abscess, hemorrhage K57.12 Diverticulitis of small intestine without perforation and abscess, no hemorrhage K57.13 Diverticulitis of small intestine without perforation and abscess, hemorrhage K57.22 Diverticulitis of large intestine with perforation and abscess, no hemorrhage K57.23 Diverticulitis of large intestine with perforation and abscess, hemorrhage K57.32 Diverticulitis of large intestine without perforation and abscess, no hemorrhage K57.33 Diverticulitis of large intestine without perforation and abscess, hemorrhage K57.42 Diverticulitis of both small and large intestine with perforation and abscess, no hemorrhage K57.43 Diverticulitis of both small and large intestine with perforation and abscess, hemorrhage K57.52 Diverticulitis of both small and large intestine without perforation and abscess, no hemorrhage K57.53 Diverticulitis of both small and large intestine without perforation and abscess, hemorrhage K57.82 Diverticulitis of intestine, part unspecified, with perforation and abscess, no hemorrhage K57.83 Diverticulitis of intestine, part unspecified, with perforation and abscess, hemorrhage K57.92 Diverticulitis of intestine, part unspecified, without perforation or abscess, no hemorrhage

K57.93 Diverticulitis of intestine, part unspecified, without perforation or abscess, hemorrhage . K61 Abscess of anal and rectal regions K65 Peritonitis K63.0 Abscess of intestine K63.1 Perforation of intestine (nontraumatic) K75.0 Abscess of liver K75.1 Phlebitis of portal vein K81.0 Acute cholecystitis N10 Acute tubulointerstitial nephritis N11 Chronic tubulointerstitial nephritis N12 Tubulointerstitial nephritis, not specified as acute or chronic N34 Urethritis and urethral syndrome N39.0 Urinary tract infection, site not specified N41 Inflammatory diseases of prostate N70 Salpingitis and oophoritis N71 Inflammatory disease of uterus, except cervix N72 Inflammatory disease of cervix uteri N73 Other female pelvic inflammatory diseases N74 Female pelvic inflammatory disorders in diseases classified elsewhere N75 Diseases of Bartholin's gland N76 Other inflammation of vagina and vulva N77 Vulvovaginal ulceration and inflammation in diseases classified elsewhere L03 Cellulitis L04 Acute lymphadenitis L08 Other local infections of skin and subcutaneous tissue L88 Pyoderma gangrenosum M00 Pyogenic arthritis M86 Osteomyelitis A49.9 Bacterial infection, unspecified T82.6 Infection and inflammatory reaction due to cardiac valve prosthesis T82.7 Infection and inflammatory reaction due to other cardiac and vascular devices, implants and grafts T83.5 Infection and inflammatory reaction due to prosthetic device, implant and graft in urinary system T83.6 Infection and inflammatory reaction due to prosthetic device, implant and graft in genital tract T84.5 Infection and inflammatory reaction due to internal joint prosthesis T84.6 Infection and inflammatory reaction due to internal fixation device T84.7 Infection and inflammatory reaction due to other internal orthopedic prosthetic devices, implants and grafts T85.7 Infection and inflammatory reaction due to other internal prosthetic devices, implants and grafts T81.4 Infection following a procedure, not elsewhere classified

T88.0 Infection following a procedure, not elsewhere classified T88.0 Infection following immunization

ICD-10-AM translation of organ dysfunction codes used by Angus et al¹³

R57 Shock, not elsewhere classified D69.6 Thrombocytopenia, unspecified 195 Hypotension D69.8 Other specified hemorrhage conditions D69.9 Hemorrhagic condition, unspecified Block 569 Continuous ventilatory support D65 Disseminated intravascular coagulation (defibrination G93.4 Encephalopathy, unspecified F05 Delirium, not induced by alcohol and other psychoactive syndrome) K72.0 Acute and subacute hepatic failure Substances G93.1 Anoxic brain damage, not elsewhere classified K76.3 Infarction of liver D69.5 Secondary thrombocytopenia N17 Acute renal failure 1. Explicit criteria (Presence of any one of the following ICD codes)

Appendix B: Modified IHME/GBD_ICD coding

method to identify sepsis

Sepsis-specific ICD-10 code

A02.1 Salmonella sepsis

A20.7 Sepsis due to plague A42.7 Sepsis due to actinomyces A21.7 Sepsis due to tularemia A22.7 Sepsis due to anthrax A24.1 Sepsis due to melioidosis A26.7 Sepsis due to erysipelothrix A28.2 Sepsis due to extraintestinal versiniosis A32.7 Sepsis due to listeria monocytogenes A39.4 Meningococcemia, unspecified A40 Streptococcal sepsis A41 Other sepsis

A54.86 Sepsis due to gonococcal infection B00.7 Disseminated herpesviral disease B37.7 Candidal sepsis R57.2 Septic Shock R65.1 Systemic Inflammatory Response Syndrome of infectious origin with organ failure O85 Puerperal sepsis

Implicit criteria (Presence of one infection code plus one organ dysfunction code)

Infection-related ICD-10 codes

A00	Cholera
A01	Typhoid fever
A02	Other salmonella infections
A03	Shigellosis
A04	Other bacterial intestinal infections
A05	Other bacterial food-borne intoxications
A06	Amebiasis
A07	Other protozoal intestinal diseases
A08	Viral and other specified intestinal infections
A09	Diarrhea and gastroenteritis, presumed infectious
origin	
A19	Miliary tuberculosis
A20	Plague
A21	Tularemia
A22	Anthrax
A23	Brucellosis
A24	Glanders and melioidosis
A25	Rat-bite fevers
A26	Erysipeloid
A27	Leptospirosis
A28	Other zoonotic bacterial diseases, not elsewhere
classified	
A30	Leprosy
A31	Infection due to other mycobacteria
A32	Listeriosis
A36	Diphtheria
A37	Whooping cough
A38	Scarlet fever
A39.0	Meningococcal meningitis
A39.1	Waterhouse-Friderichsen syndrome
A39.2	Acute meningococcemia
A39.3	Chronic meningococcemia
A39.4	Meningococcemia, unspecified
A39.5	Meningococcal heart disease
A39.81	Meningococcal encephalitis
A39.82	Meningococcal retrobulbar neuritis
A39.89	Other meningococcal infections
A39.9	Meningococcal infection, unspecified
A42	Actinomycosis
A43	Nocardiosis
A44	Bartonellosis
A46	Erysipelas
A48	Other bacterial diseases, elsewhere classified
A48.0	Gas gangrene
A48.1	Legionnaires disease
A48.3	Toxic shock syndrome
A49	Bacterial infection of unspecified site
A49.9	Bacterial infection, unspecified
A54	Gonococcal infection
A59	Trichomoniasis
A69.0	Necrotising ulcerative stomatitis
A69.1	Other Vincent's infections
A69.9	Spirochetal infection, unspecified
A70	Chlamydia psittaci infections
A74	Other diseases caused by chlamydiae
	The last factors in

~/+	Other diseases ta
A75	Typhus fever

A77	Spotted fever (tick-borne rickettsioses)
A78	Q fever
A79	Other rickettsioses
A80	Acute poliomyelitis
A81	Atypical virus infections of CNS
A83	Mosquito-borne viral encephalitis
A84	Tick-borne viral encephalitis
A85	Other viral encephalitis, not elsewhere classified
A86	Unspecified viral encephalitis
A87	Viral meningitis
A88	Other viral infections of CNS, not elsewhere classifie
A89	Unspecified viral infection of CNS
A90	Dengue fever
A91	Dengue hemorrhagic fever
A92	Other mosquito-borne viral fevers
A93	Other arthropod-borne viral fevers, not elsewhe
classified	
A94	Unspecified arthropod-borne viral fever
A95	Yellow fever
A96	Arenaviral hemorrhagic fever
A98	Other viral hemorrhagic fever, not elsewhe
classified	
A99	Unspecified viral hemorrhagic fever
B00	Herpesviral [herpes simplex] infections
B01	Varicella
B02	Zoster
B03	Smallpox
B04	Moneypox
B05	Measles
B06	Rubella
B08	Other viral infections characterized by skin ar
mucous m	nembrane lesions, not elsewhere classified
B09	Unspecified viral infection characterized by skin an
mucous m	nembrane lesions
B10	Other human herpesviruses
B25	Cytomegaloviral disease
B26	Mumps
B27	Infectious mononucleosis
B33	Other viral diseases, not elsewhere classified
B34	Viral infection of unspecified site
B37	Candidiasis
B38	Coccidioidomycosis
B38.7	Disseminated coccidioidomycosis
B39	Histoplasmosis
B39.3	Disseminated histoplasmosis capsulati
B40	Blastomycosis
B40.7	Disseminated blastomycosis
B41	Paracoccidioidomycosis
B41.7	Disseminated paracoccidioidomycosis
	Sporotrichosis
B42	
B42 B42.7	Disseminated sporotrichosis
	•
B42.7	Disseminated sporotrichosis
B42.7 B43	Disseminated sporotrichosis Chromomycosis and phaeomycotic abscess Aspergillosis
B42.7 B43 B44	Disseminated sporotrichosis Chromomycosis and phaeomycotic abscess

B46	Zygomycosis					
B46.4	Disseminated mucormycosis					
B48	Other mycoses, not elsewhere classified					
B49	Unspecified mycosis					
B50	Pasmodium falciparum malaria					
B50.8	Other severe and complicated Plasmodium					
falciparum						
B54	Unspecified malaria					
B55	Leishmaniasis					
B57	Chagas Disease					
B58 B59	Toxoplasmosis Pneumocystosis					
B60	Other protozoal diseases, not elsewhere classified					
B64	Unspecified protozoal disease					
B67	Echinococcosis					
B95	Streptococcus, Staphylococcus, and Enterococcus as					
the cause of	of diseases classified elsewhere					
B96	Other bacterial agents as the cause of diseases					
classified e	lsewhere					
B97	Viral agents as the cause of diseases classified					
elsewhere						
B99	Other and unspecified infectious diseases					
G00	Bacterial meningitis, not elsewhere classified					
G01	Meningitis in bacterial diseases classified elsewhere					
G02	Meningitis in other infectious and parasitic diseases					
classified e						
G03	Meningitis due to other and unspecified causes					
G04 G05	Encephalitis, myelitis and encephalomyelitis					
	Encephalitis, myelitis and encephalomyelitis in assified elsewhere					
G06	Intracranial and intraspinal abscess and granuloma					
G07	Intracranial and intraspinal abscess and granuloma in					
	assified elsewhere					
G08	Intracranial and intraspinal phlebitis and					
thromboph	· ·					
H05.01	Cellulitis of orbit					
H05.02	Osteomyelitis of orbit					
H05.03	Periostitis of orbit					
H60.2	Malignant otitis externa					
H70.0	Acute mastoiditis					
100	Rheumatic fever without heart involvement					
126.01	Septic pulmonary embolsim with acute cor					
pulmonale	Contin automatic controline without contrology					
126.90	Septic pulmonary embolism without acute cor					
pulmonale 133	Acute and subacute endocarditis					
133	Endocarditis, valve unspecified					
139	Endocarditis and heart valve disorders in diseases					
classified e						
140.0	Infective myocarditis					
176	Septic non-pulmonary arterial embolism					
196	Gangrene, not elsewhere classified					
J01	Acute sinusitis					
J02	Acute pharyngitis					
J03	Acute tonsillitis					
J04	Acute laryngitis and tracheitis					
J05	Acute obstructive laryngitis (croup) and epiglottitis					
J06	Acute upper respiratory infections of multiple and					
unspecified						
J09	Influenza due to certain identified influenza viruses					
J10	Influenza due to other identified influenza virus					
J11	Influenza due to unidentified influenza virus					
J12	Viral pneumonia, not elsewhere classified					
J13 J14	Pneumonia due to Streptococcus pneumoniae Pneumonia due to Haemophilus influenzae					
J14 J15	Bacterial pneumonia, not elsewhere classified					
J15 J16	Pneumonia due to other infectious organisms, not					
elsewhere	-					
J17	Pneumonia in diseases classified elsewhere					
J18						
110	Pneumonia, organism unspecified					

J21	Acute bronchiolitis				
J22	Unspecified acute lower respiratory infection				
J36	Peritonsillar abscess				
J39.0 J39.1	Retropharyngeal and parapharyngeal abscess Other abscess of pharynx				
J85	Abscess of lung and mediastinum				
J86	Pyothorax				
K35	Acute appendicitis				
K36	Other appendicitis				
K37	Unspecified appendicitis				
K57	Diverticulitis disease of intestine				
K61 K63.0	Abscess of anal and rectal regions Abscess of intestine				
K63.0	Perforation of intestine (nontraumatic)				
K65	Peritonitis				
K75.0	Abscess of liver				
K81.0	Acute cholecystitis				
K81.2	Acute cholecystitis with chronic cholecystitis				
K83.0	Cholangitis				
K95.01 K95.81	Infection due to gastric band procedure Infection due to other bariatric procedure				
L02	Cutaneous abscess, furuncle and carbuncle				
L02	Cellulitis				
L04	Acute lymphadenitis				
L08	Other local infections of skin and subcutaneous tissue				
M00	Pyogenic arthritis				
M01	Direction infections of joint in infectious and parasitic				
diseases c M86	lassified elsewhere				
N10	Osteomyelitis Acute pyelonephritis				
N15.1	Renal and perinephric abscess				
N30	cystitis				
N39.0	Urinary tract infection, site not specified				
N41.0	Acute prostatitis				
N41.2	Abscess of prostate				
N41.3	Prostatocystitis Orchitic and anididumitic				
N45 N70	Orchitis and epididymitis Salpingitis and oophoritis				
N71	Inflammatory disease of uterus, except cervix				
N72	Inflammatory disease of cervix uteri				
N73	Other female pelvic inflammatory diseases				
N74	Female pelvic inflammatory disorders in diseases				
classified e					
N98.0 O03.0	Infection associated with artificial insemination Genital tract and pelvic infection following				
	e spontaneous abortion				
003.5	Genital tract and pelvic infection following complete				
or unspec	ified spontaneous abortion				
003.88	Urinary tract infection following complete or				
-	d spontaneous abortion				
004.5	Genital tract and pelvic infection following (induced)				
004.88	on of pregnancy Urinary tract infection following (induced)				
	on of pregnancy				
007.38	Urinary tract infection following failed attempted				
terminatio	on of pregnancy				
008.0	Genital tract and pelvic infection following ectopic				
	pregnancy				
008.83	Urinary tract infection following ectopic and molar				
pregnancy O23	Infections of genitourinary tract in pregnancy				
075.3	Other infection during labour				
086	Other puerperal infections				
088.3	Obstetric pyemic and septic embolism				
091	Infections of breast associated with pregnancy, the				
	m and lactation				
098 elsewberg	Maternal infectious and parasitic diseases classifiable but complicating pregnancy, childbirth and the				
puerperiu					
R78.81	Bacteraemia NOS				

T80.2 Sepsis following infusion, transfusion, or therapeutic injection

T81.4 Infection following a procedure, not elsewhere classified

T82.6 Infection and inflammatory reaction due to cardiac valve prosthesis

T82.7 Infection and inflammatory reaction due to other cardiac and vascular devices, implants and grafts

T83.5 Infection and inflammatory reaction due to prosthetic device, implant and graft in urinary system

T83.6 Infection and inflammatory reaction due to prosthetic device, implant and graft in genital tract

T84.5 Infection and inflammatory reaction due to internal joint prosthesis

T84.6 Infection and inflammatory reaction due to internal fixation device

T84.7 Infection and inflammatory reaction due to other internal orthopedic prosthetic devices, implants and grafts

T85.7 Infection and inflammatory reaction due to other internal prosthetic devices, implants and grafts

T88.0 Sepsis following immunization

U04 Severe Acute Respiratory Syndrome (SARS)

2. Organ Dysfunction Codes

D65	Disseminated	intravascular	coagulation		
[defibrina	tion syndrome]				
D69.5	Secondary thrombocytopenia			K72.9	Hepatic failure, unspecified
E87.2	Acidosis			N00	Acute nephritic syndrome
G93.4	4 Other and unspecified encephalopathy			N17	Acute kidney failure
146	Cardiac arrest			R09.02	Hypoxemia
195.9	Hypotension, uns	specified		R09.2	Respiratory arrest
J80	Acute respiratory distress syndrome			R40.0	Somnolence
J95.2	Acute pulmonary insufficiency following nonthoracic		R40.1	Stupor	
surgery				R40.2	Coma
J96	Respiratory failur	re, not elsewhere cla	ssified	R40.20	Unspecified coma
J96.0	Acute respiratory	/ failure		R41.82	Altered mental status, unspecified
J96.2	Acute and chronic respiratory failure			R55	Syncope and collapse
J96.9	Respiratory failur	e, unspecified		R57	Shock, not elsewhere classified
K72.0	Acute and subacu	ute hepatic failure			

Appendix C: SOFA calculation worksheet

ORGAN SYSTEM	0 1 2 3 4		4	9	Organ				
ORGAN SYSTEM	U	1	2	3	4	9	scores		
Respiration									
PaO ₂ / FIO ₂									
(in mmHg)	>400	301 - 400	201 – 300 (with respiratory support*) <301 (without respiratory support*)	101 - 200 ≤ 100 (with respiratory support*) (with respiratory support*)		Variable not			
(in kPa)	>53.2	40.0 - 53.1	26.7 – 39.9 (with respiratory support*) <40.0 (without respiratory support*)	13.4 – 26.6 (with respiratory support*)	≤ 13.3 (with respiratory support*)	measured			
<u>Cardiovascular</u>									
Hypotension	MAP > 70 mmHg	MAP < 70 mmHg	dopamine ≤ 5.0 (doses are given in µg / kg / minute) or any dose dobutamine or any dose milrinone or any dose levosimendan	dopamine 5.0 – 15.0 (doses are given in $\mu g / kg / minute)$ or epinephrine ≤ 0.1 or norepinephrine ≤ 0.1 or any dose vasopressin or any dose metaraminol or any dose phenylephrine	dopamine > 15.0 (doses are given in μg / kg / minute) or epinephrine >0.1 or norepinephrine >0.1	Variable not measured			
Coagulation									
Platelets (x 10 ⁹ / L)	>150	101 - 150	51 - 100	21 - 50	≤ 20	Variable not measured			
<u>Liver</u>									
Bilirubin									
(mg / dl)	< 1.2	1.2 - 1.9	2.0 - 5.9	6.0 - 11.9	> 12.0	Variable not			
(µmol / L)	<20	20 - 32	33 - 101	102 - 204	>204	measured			
Renal									
Creatinine			1	1					
(mg / di)	< 1.2	1.2 - 1.9	2.0-3.4	3.5 - 4.9	> 5.0	Variable not			
(µmol/I)	< 110	110- 170	171 - 299	300 - 440	> 440	measured			
OR Urine output				or < 500 ml / day	or < 200 ml / day				
Central Nervous System (GCS)									
	15	13-14	10-12	6-9	<6				